

Salivary Hormone Testing

Policy Number: AHS – G2120 – Salivary Hormone Testing	Prior Policy Name and Number, as applicable: G2120 – Salivary Hormone Testing for Menopause and Aging
Initial Presentation Date: 09/18/2015 Revision Date: 03/01/2023	

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I. Policy Description

Testing of saliva has been proposed as a noninvasive method to measure free (unbound to carrier proteins) steroid hormones, including estrogen, progesterone, androgens, and cortisol, for diagnosis of hormonal imbalance and administration of individualized hormone replacement therapy (ACOG & ASRM, 2012).

Hypercortisolism can occur in several disorders, including Cushing syndrome (pituitary hypersecretion of corticotropin/ACTH), or glucocorticoid administration resulting in obesity, hypertension, menstrual irregularity, and glucose intolerance (Lacroix et al., 2015; Nieman et al., 2008; Nieman, 2022a; Quddusi et al., 1998).

Terms such as male and female are used when necessary to refer to sex assigned at birth.

II. Indications and/or Limitations of Coverage

Application of coverage criteria is dependent upon an individual's benefit coverage at the time of the request. Specifications pertaining to Medicare and Medicaid can be found in [Applicable State and Federal Regulations](#) of this policy document."

- 1) For the diagnosis of Cushing syndrome, late night salivary cortisol testing **MEETS COVERAGE CRITERIA**.

The following does not meet coverage criteria due to a lack of available published scientific literature confirming that the test(s) is/are required and beneficial for the diagnosis and treatment of a patient's illness.

- 2) For the screening, diagnosis, **and/or** monitoring of menopause, infertility, endometriosis, polycystic ovary disease (PCOS), premenstrual syndrome, osteoporosis, sexual dysfunction, seasonal affective disorder, depression, multiple sclerosis, sleep disorders, **or** diseases related to aging, salivary hormone testing **DOES NOT MEET COVERAGE CRITERIA**.

III. Table of Terminology

Term	Definition
AACE	American Association of Clinical Endocrinologists

Term	Definition
ACOG	American College of Obstetricians and Gynecologists
ACTH	Adrenocorticotrophic hormone
ASRM	American Society of Reproductive Medicine Practice Committee
CLIA '88	Clinical Laboratory Improvement Amendments of 1988
CMS	Centers for Medicare and Medicaid
CS	Cushing syndrome
DHEA	Dehydroepiandrosteron
E1	Estrone
E2	Estradiol
E3	Estriol
EIA	Enzyme immunoassay
ELISA	Enzyme-linked immunosorbent assay
ERCUSYN	European Registry on Cushing's Syndrome
ES	Endocrine Society
FDA	Food and Drug Administration
HT	Hormone therapy
IVF	In vitro fertilization
LC-MS	Liquid chromatography with tandem mass spectrometry
LDTs	Laboratory-developed tests
LNCS	Late night salivary cortisol
MHT	Menopausal hormone therapy
MP	Micronized progesterone
MS	Multiple sclerosis
NAMS	North American Menopausal Society
PCOS	Polycystic ovary disease
RIA	Radioimmunoassay
TSS	Transsphenoidal selective adenectomy

IV. Scientific Background

Testing of hormone levels in the saliva has been proposed as a noninvasive method to measure free (unbound to carrier proteins and thus active) steroid hormones (estrogen, progesterone, androgens, cortisol, etc.) for diagnosis of hormonal imbalance and administration of individualized hormone replacement therapy (ACOG & ASRM, 2012). Saliva measurements are thought to represent the concentrations of unconjugated steroid hormones as well as unconjugated steroids that have diffused freely into saliva. Conjugated steroids will often show significant decreases in concentration because their filtration process into the saliva is limited. This is what causes hormones, such as cortisol, estradiol, and testosterone to approximate concentrations well and the hormone dehydroepiandrosterone (DHEA) to represent concentrations poorly (Wood, 2009).

Salivary hormone level testing is often recommended by bioidentical hormone vendors as a means of providing personalized therapy. However, individualized testing and monitoring is only

useful when a narrow therapeutic window exists for a drug or a drug class. Steroid hormones, such as estrogen and progesterone, do not meet these criteria and do not require individualized testing (ACOG & ASRM, 2012; Conaway, 2011). Furthermore, there is no evidence that hormonal levels in saliva are biologically meaningful. Saliva is an ultra-filtrate of the blood and in theory, should be amenable to testing for free concentrations of hormones; however, salivary testing does not appear to be an accurate or precise method of hormone testing (Flyckt et al., 2009; Lewis et al., 2002). Studies suggest that salivary assessments of hormone levels are inaccurate and do not correlate with levels determined from serum (Conaway, 2011), as there is large within-patient variability in salivary hormone concentrations, especially when exogenously administered hormones are given (Hardiman et al., 1990; Klee & Hesser, 2000; Lewis et al., 2002; Meulenberget al., 1987; Wren et al., 2000). Salivary hormone levels often fluctuate with factors, such as circadian rhythm, and frequently do not correlate well with serum levels of hormones (Wood, 2009).

Salivary hormone measurement may be utilized for many purposes. Menopause occurs due to changing hormone levels, mainly estrogen. In general, [individuals] experience menopause at a mean age of 51 years, with most becoming menopausal between 45 and 55. Menopausal hormone therapy (MHT, estrogen alone or combined with a progestin) is used for management of menopausal symptoms and is highly effective for symptoms, such as hot flashes and vaginal atrophy. In some cases, MHT may be used for the mood lability that many [individuals] experience during the menopausal transition (Martin & Barbieri, 2022; Taylor & Manson, 2011). There are few indications for the measurement of hormone levels to evaluate success of therapy when treating a postmenopausal [individual] with hormones. If treatment is initiated for symptom control, therapy should be titrated to the alleviation of symptoms, not a laboratory value (ACOG & ASRM, 2012). A salivary hormone test has been developed by Genova Diagnostics, which evaluates levels of hormones in [individuals] during perimenopause, menopause, and andropause (Genova_Diagnostics, 2020).

One of the primary hormones that diffuses freely into saliva and can be well-approximated by salivary measurements is cortisol. Cortisol is a steroid hormone that is produced due to stress. Salivary flow rate does not affect cortisol concentration, and salivary cortisol correlates well with serum-free cortisol. This property can be used to identify adrenal insufficiencies and other related disorders (Nieman, 2019). For example, the presence of Cushing syndrome (CS) is suggested by signs of hypercortisolism, such as proximal myopathy, facial plethora, and wide purplish striae. However, none of these are pathognomonic, and many are nonspecific (such as obesity or hypertension). As a result, the diagnosis must be confirmed by biochemical tests, one of which is a salivary cortisol measurement (Nieman, 2022b). The recurrence of hypercortisolemia after an initial treatment for CS seems to be predicted earlier by late night salivary cortisol (LNSC) testing compared to urinary free cortisol excretion (Fleseriu et al., 2016).

Proprietary Testing

Saliva testing measures the amount of hormone available to target tissues and is a good option for monitoring hormonal therapy. ZRT Laboratories developed a Saliva Steroid Profile using liquid chromatography/tandem mass spectrometry (LC-MS/MS) which tests a broad range of bioavailable hormones and hormone metabolites in one saliva sample collection. LC-MS/MS testing accurately reports levels of estrogen, such as those seen in men, children, and people using aromatase inhibitors, and includes a test for ethinyl estradiol, three hormone blockers, and melatonin. "Testing the levels of both upstream precursors and downstream metabolites of these parent active steroids [estrogens, progestogens, androgens, glucocorticoids, mineralocorticoids, melatonin, synthetic estrogen ethinyl, estradiol, anastrozole, letrozole, and

the 5 α -reductase inhibitor finasteride] will help determine which steroid synthesis enzymes are low, overactive, blocked by natural or pharmaceutical inhibitors, or defective due to metabolic dysfunctions (e.g., Polycystic Ovarian Syndrome (PCOS), Premenstrual Dysphoric Disorder (PMDD), luteal dysfunction, overexpression of aromatase, and estrogen dominance) and inborn errors of metabolism such as Congenital Adrenal Hyperplasia (CAH)” (ZRTLAB, 2019). ZRT is one of the first labs to measure hormones in saliva and helped establish the method that made saliva hormone testing commercially viable for health care providers and patients around the globe.

UnikeyHealth developed a saliva hormone testing panel to assess six hormone levels with an at-home test. The hormones tested are progesterone, estradiol, estriol, testosterone, DHEA, and cortisol. This at-home test provides recommendations and is purported to identify underlying causes of hormonal imbalance issues based on the individualized hormone assessment (UnikeyHealth, 2022).

Genova Diagnostics has developed several saliva hormone tests including The Rhythm™ hormone test (Genova, 2022d), Menopause Plus™ (Genova, 2022c), The Comprehensive Melatonin Profile (Genova, 2022b), and The Adrenocortex Stress Profile (Genova, 2022a). The Rhythm™ hormone test is a comprehensive assessment of estradiol, progesterone, and testosterone which can help assess underlying causes of disorders such as premenstrual syndrome (PMS), infertility, and menstrual irregularities (Genova, 2022d). Menopause Plus™ is Genova's most comprehensive salivary hormone profile and is designed to provide insight into the impact that shifting hormone levels may play in men (andropause or male menopause) and women (peri/menopause). This test collects eight saliva samples every other day over six days for estrone (E1), estradiol (E2), estriol (E3), progesterone, progesterone/estradiol ratio (P/E2), and testosterone (Genova, 2022c). The Comprehensive Melatonin Profile analyzes the circadian secretion patterns of melatonin by analyzing three saliva samples taken in the morning, afternoon, and midnight. This test is purported to determine underlying causes of melatonin imbalances in sleep disorders, depression, and seasonal affective disorder (Genova, 2022b). Lastly, The Adrenocortex Stress Profile (ASP) provides an assessment of the Hypothalamic-Pituitary-Adrenal (HPA) axis using carefully timed salivary samples of cortisol and DHEA. This may help reveal HPA axis imbalances which could be a contributing factor in cardiovascular disease, immune dysregulation, diabetes, chronic fatigue, persistent pain, or cognitive decline (Genova, 2022a).

Analytical Validity

Multiple proprietary tests are available for salivary hormone testing. Tests such as ZRT and UnikeyHealth ask the user to submit saliva samples and send the specimen to the proprietary lab where it can be analyzed. Labs will typically use an immunoassay-based method, such as an enzyme-linked immunosorbent assay (ELISA) or enzyme immunoassay (EIA), to assess the concentration of hormones, such as estradiol or progesterone. Others may use an automated competitive electrochemiluminescence immunoassay for LNSC measurement (Spence et al., 2018). The results are compiled into a report listing the concentrations of each hormone as well as comments on abnormal amounts. These tests are often marketed to post-menopausal [individuals] who desire to have an assessment of hormones like estrogen, progesterone, DHEA, testosterone, estriol, and cortisol (UnikeyHealth, 2022; ZRTLAB, 2019). Moreover, another proprietary test proposes that conditions such as multiple sclerosis (MS) can be assessed through irregularities in melatonin (Genova, 2022b). However, not only is melatonin not widely measured through saliva, but there is currently no compelling data for whether administering melatonin has any utility with dealing with MS; there has been far too little

published data with human subjects to draw any conclusions (Wurtman, 2017). Osteoporosis is another condition that tests may purportedly be able to screen for with saliva (Genova, 2022d). However, this test may be of limited utility as the risks of hormone therapy may outweigh the benefits (Rossouw et al., 2002).

Salivary cortisol was first measured by direct radioimmunoassay (RIA) in 1978, but more accurate cortisol immunoassays have now been developed; however, these assays are often limited due to poor specificity (El-Farhan et al., 2017). Further, late at night, cortisol levels may fall below detection limits for some RIA testing methods. Liquid chromatography with tandem mass spectrometry (LC-MS/MS) has also been used for the detection of salivary cortisol. Schiffer et al. (2019) developed a novel LC-MS/MS assay to identify androgens in saliva samples with appropriate sensitivity. Prior, Li et al. (2018) was able to utilize the same technique (LC-MS/MS) to accurately quantify three estrogens (estrone E1, estradiol E2, and estriol E3) in an assay with an accuracy of 98.9-112.4% and precision of ($\leq 7.4\%$) as a hopeful alternative to blood samples. However, this field continues to face limitations due to poorly standardized assays and a lack of a single, validated reference range (El-Farhan et al., 2017).

Initial diagnostic tests for hypercortisolism should be highly sensitive, even if the diagnosis may be excluded later. Late night salivary cortisol (LNSC) is a first-line diagnostic test for CS as indicated by the approach outlined by the 2008 Endocrine Society (Nieman et al., 2008) and others (Hinojosa-Amaya et al., 2019). LNSC measurements are obtained at least twice because the hypercortisolism in CS may be variable. Two measurements must be abnormal for the test to be considered abnormal; this may be especially difficult for patients with fluctuating disease. The diagnosis of CS is established when at least two different first-line tests (such as LNSC and 24-hour urinary cortisol excretion) are abnormal. Once the diagnosis is established, additional evaluation is done to identify the cause of the hypercortisolism (Nieman, 2022b).

A locally modified RIA assay was developed by Nunes et al. (2009) and measured LNSC in obese patients with a current or past diagnosis of CS. The assay was able to diagnose a recurrence of CS with a sensitivity of 90% and a specificity of 91.8%; it was also reported that "A threshold of 12 nmol/liter yielded 100% sensitivity and specificity in overt [Cushing] syndrome" (Nunes et al., 2009).

Ueland et al. (2021) studied the analytical validity of late-night salivary cortisol as a screening test for CS. Bedtime and morning salivary samples were collected from 54 children in the obesity clinic and three children with pituitary CS using liquid chromatography tandem mass spectrometry (LC-MS/MS). These levels were compared to 320 salivary samples from healthy children to set cut-off values. Bedtime cutoff levels for cortisol and cortisone were 2.4 and 12.0 nmol/L, respectively. "Applying these cutoff levels on the verification cohort, 1 child from the obesity clinic had bedtime salivary cortisol exceeding the defined cutoff level, but normal salivary cortisone. All 3 children with pituitary CS had salivary cortisol and cortisone far above the defined bedtime cutoff levels. Healthy subjects showed a significant decrease in salivary cortisol from early morning to bedtime" (Ueland et al., 2021). The authors conclude that bedtime salivary cortisol levels with a diagnostic threshold above 2.4 nmol/L can be applied as a screening test for CS in children.

Clinical Utility and Validity

A study by Lewis et al. (2002) focusing on salivary progesterone measurements found major variation when a progesterone cream was applied to several post-menopausal [individuals]. Salivary measurements were collected at zero, one, three, four, seven, and eight weeks. The

average baseline for the 20 mg/g cream group was found to be 0.25 ± 0.12 nmol/L, but the measurement at one week was 82.11 ± 104.52 nmol/L (Lewis et al., 2002); similar enormous variations were found at three and seven weeks, as well as the 40 mg/gm cream group. In contrast, the placebo group's baseline was 0.43 ± 0.21 and 0.38 ± 0.20 in week eight (Lewis et al., 2002). The finding with inconsistent salivary progesterone levels was even found among premenopausal [individuals] obtaining *in vitro* fertilization (IVF); on the other hand, salivary estradiol was found to be correlative to serum-based assessment, and could be a less invasive alternative to blood draws for ovarian stimulation during IVF cycles (Sakkas et al., 2020).

LNSC measurements were found to be concordant with the 24-hour urine test, with 97% concordance at ≥ 4 nmol/L and 69% concordance at ≥ 10 nmol/L. However, the tests were stated to be "equivalent" at the more sensitive cutoff of 4 nmol/L. The authors concluded that due to the concordance of the salivary test with the urine test, the salivary test should replace the urinary test as the frontline test for Cushing syndrome (Doi et al., 2013). Another study found LNSC to be 100% sensitive and 98% specific at a cut-off of 2.4 nmol/L (Antonelli et al., 2015). Both cortisol and its metabolite cortisone were tested as cortisone is a significant source of interference in certain immunoassays. The variation between and within runs were both under ten percent, the method was linear up to 55.4 nmol/L for cortisol, and the lower limit of quantification was 0.51 nmol/L for cortisol (Antonelli et al., 2015).

A study measured the utility of salivary testosterone and cortisol concentrations in 71 junior athletes (26 females and 45 males) in response to stress. The researchers compared results of salivary samples to capillary blood samples taken at the same time; while blood samples showed an increase in both testosterone and cortisol concentrations in both sexes, salivary samples showed no change in testosterone or cortisol levels (Crewther et al., 2018). This may suggest that salivary hormone testing in these populations is not as efficient as other methods.

Valassi et al. (2017) analyzed diagnostic data from 1341 CS patients in the European Registry on Cushing's syndrome (ERCUSYN) and noted that of the three main first-line CS diagnostic tests, the urinary free cortisol test was performed in 78% of patients as a first-line testing method, overnight 1 mg dexamethasone suppression test was performed in 60% of patients, and LNSC was performed in only 25% of patients. This shows that LNSC may not be used as frequently as other testing methods for a first-line diagnosis of CS.

Salivary testing for cortisol could also prove useful in occupational settings as a parameter for stress. Oldenburg and Jensen (2019) conducted a study on merchant ship crew, and found that after adjustment, average salivary cortisol level was positively associated with "acute shipboard stressors, namely the average current working time ($p=.050$) and the average number of terminals that had been served during the last 7 days ($p=0.008$)."

This laboratory data is essential in all fields wherein professionals experience high levels of stress, so that measures can be taken to create a positive working environment.

Kim et al. (2020) studied the diagnostic utility of stimulated salivary cortisol as a noninvasive diagnostic tool for adrenal insufficiency (AI). One hundred twenty subjects were measured for stimulated cortisol levels and these levels were compared to those obtained from the short Synacthen test (SST). AI was defined as a cortisol level of <496.8 nmol/L during the SST. Thirty-four of 120 patients were diagnosed with AI according to SST results. "Basal and stimulated salivary cortisol levels were positively correlated with basal ($r=0.538$) and stimulated serum cortisol levels ($r=0.750$), respectively (all $P<0.001$)."

The cut off level of morning basal salivary cortisol was 3.2 nmol/L, and the cutoff value of stimulated salivary cortisol was 13.2 nmol/L.

Subjects with a stimulated salivary cortisol level above 13.2 nmol/L but a stimulated serum cortisol level below 496.8 nmol/L (n=2) had lower serum albumin levels than those showing a concordant response. The authors conclude that "The diagnostic performance of stimulated salivary cortisol measurements after the SST was comparable to serum cortisol measurements for diagnosing AI" (Kim et al., 2020).

V. Guidelines and Recommendations

American Association of Clinical Endocrinologists (AACE)

The AACE has noted salivary hormone level testing as recommended by certain proponents to provide individualized therapy. However, these methods are not FDA or CLIA approved, and factors such as hydration and circadian rhythm may influence the concentration of hormones within a subject. Standardization is difficult, and even though standardized blood tests do exist; it is of limited clinical utility because measuring hormone levels in postmenopausal [individuals] has no predictive value on what the normal levels should be. A salivary measurement cannot be used to correct the levels of sex hormones (Goodman et al., 2011).

American College of Obstetricians and Gynecologists (ACOG) and the American Society of Reproductive Medicine Practice Committee (ASRM)

ACOG and ASRM released joint guidelines on compounded hormone therapy that stated salivary hormone testing had no evidence to support its biological utility and that testing the hormone levels were neither accurate nor precise. The guidelines stated that salivary hormone testing had large intra-patient variability depending on factors such as diet and that saliva did not provide a reasonable representation of serum hormone levels. Saliva may be contaminated with other cell types, contains lower concentration of hormones than serum, and impossible to reliably test for a representative result. The guidelines concluded that evidence is inadequate to support an individualized hormone therapy based on salivary, serum, or urine testing (ACOG & ASRM, 2012).

Finally, the guideline wrote that "there is no evidence that hormonal levels in saliva are biologically meaningful. In addition, whereas saliva is an ultrafiltrate of the blood and in theory should be amenable to testing for "free" (unbound) concentrations of hormones, salivary testing does not currently offer an accurate or precise method of hormone testing " (ACOG & ASRM, 2012). This guideline was reaffirmed in 2020.

North American Menopausal Society (NAMS)

The NAMS addressed salivary hormone testing with regards to MHT, stating that salivary hormone testing is "inaccurate and unreliable." The NAMS further notes that the levels in serum, saliva, and tissue are "markedly different" and alludes to the FDA's statement that there is "no scientific basis for using saliva testing to adjust hormone levels" (NAMS, 2012).

The NAMS also addressed salivary hormone testing in the context of compounded HT (hormone therapy), which would include estradiol, estrone, and micronized progesterone (MP), but corroborates that salivary testing for HT is considered "unreliable because of differences in hormone pharmacokinetics and absorption, diurnal variation, and inter-individual and intraindividual variability" (NAMS, 2017).

Endocrine Society (ES)

The ES states that “salivary hormone assays are not standardized, do not have independent quality control programs, and lack an accepted reference range.” The Society further mentions that there is no scientific evidence that a correlation exists between symptoms and salivary hormones. Assessment or monitoring of hormone therapy lacks evidence, and the American College of Obstetricians and Gynecologists, the North American Menopausal Society, and the Endocrine Society all recommend against salivary hormone testing to assess or monitor hormone levels because “they lack a rationale and therefore lead to unnecessary expense of treatment” (Santoro et al., 2016).

The ES also recommends a test of at least two LNSC measurements for diagnosis of Cushing Syndrome. If a patient has eucortisolism after a transsphenoidal selective adenomectomy (TSS), a measurement of late-night salivary or serum cortisol is recommended (Nieman, 2015; Nieman et al., 2008).

VI. Applicable State and Federal Regulations

DISCLAIMER: If there is a conflict between this Policy and any relevant, applicable government policy for a particular member [e.g., Local Coverage Determinations (LCDs) or National Coverage Determinations (NCDs) for Medicare and/or state coverage for Medicaid], then the government policy will be used to make the determination. For the most up-to-date Medicare policies and coverage, please visit the Medicare search website: <http://www.cms.gov/medicare-coverage-database/search.aspx>. For the most up-to-date Medicaid policies and coverage, visit the applicable state Medicaid website.

Food and Drug Administration (FDA)

Salivary hormones may be measured by multiple tests. Additionally, many labs have developed specific tests that they must validate and perform in house. These laboratory-developed tests (LDTs) are regulated by the Centers for Medicare and Medicaid (CMS) as high-complexity tests under the Clinical Laboratory Improvement Amendments of 1988 (CLIA '88). LDT's are not approved or cleared by the U. S. Food and Drug Administration; however, FDA clearance or approval is not currently required for clinical use.

VII. Applicable CPT/HCPCS Procedure Codes

CPT	Code Description
82530	Cortisol free
82533	Cortisol; total
82626	Dehydroepiandrosterone (dhea)
82627	Dehydroepiandrosterone
82670	Estradiol; total
82671	Estrogens; fractionated
82672	Estrogens; total
82677	Estriol
82679	Estrone
82681	Estradiol; free, direct measurement (eg, equilibrium dialysis)

CPT	Code Description
84144	Progesterone
84402	Testosterone; free
84403	Testosterone; total
84410	Testosterone; bioavailable, direct measurement (eg, differential precipitation)
S3650	Saliva test, hormone level; during menopause

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Procedure codes appearing in policy documents are included only as a general reference tool for each policy. They may not be all-inclusive.

VIII. Evidence-based Scientific References

ACOG, & ASRM. (2012). Compounded bioidentical menopausal hormone therapy. *Fertil Steril*, 98(2), 308-312. <https://doi.org/10.1016/j.fertnstert.2012.06.002>

Antonelli, G., Ceccato, F., Artusi, C., Marinova, M., & Plebani, M. (2015). Salivary cortisol and cortisone by LC-MS/MS: validation, reference intervals and diagnostic accuracy in Cushing's syndrome. *Clin Chim Acta*, 451(Pt B), 247-251. <https://doi.org/10.1016/j.cca.2015.10.004>

Conaway, E. (2011). Bioidentical hormones: an evidence-based review for primary care providers. *J Am Osteopath Assoc*, 111(3), 153-164. <https://pubmed.ncbi.nlm.nih.gov/21464264/>

Crewther, B. T., Obminski, Z., Orysiak, J., & Al-Dujaili, E. A. S. (2018). The utility of salivary testosterone and cortisol concentration measures for assessing the stress responses of junior athletes during a sporting competition. *J Clin Lab Anal*, 32(1). <https://doi.org/10.1002/jcla.22197>

Doi, S. A., Clark, J., & Russell, A. W. (2013). Concordance of the late night salivary cortisol in patients with Cushing's syndrome and elevated urine-free cortisol. *Endocrine*, 43(2), 327-333. <https://doi.org/10.1007/s12020-012-9855-0>

El-Farhan, N., Rees, D. A., & Evans, C. (2017). Measuring cortisol in serum, urine and saliva - are our assays good enough? *Ann Clin Biochem*, 54(3), 308-322. <https://doi.org/10.1177/0004563216687335>

Fleseriu, M., Hamrahian, A. H., Hoffman, A. R., Kelly, D. F., & Katznelson, L. (2016). AMERICAN ASSOCIATION OF CLINICAL ENDOCRINOLOGISTS AND AMERICAN COLLEGE OF ENDOCRINOLOGY DISEASE STATE CLINICAL REVIEW: DIAGNOSIS OF RECURRENCE IN CUSHING DISEASE. *Endocr Pract*, 22(12), 1436-1448. <https://doi.org/10.4158/ep161512.Dscr>

Flyckt, R. L., Liu, J., Frasure, H., Wekselman, K., Buch, A., & Kingsberg, S. A. (2009). Comparison of salivary versus serum testosterone levels in postmenopausal women receiving transdermal testosterone supplementation versus placebo. *Menopause*, 16(4), 680-688. <https://doi.org/10.1097/gme.0b013e318199d5c4>

Genova. (2022a). Adrenocortex Stress Profile. <https://www.gdx.net/product/adrenocortex-stress-hormone-test-saliva>

Genova. (2022b). Comprehensive Melatonin Profile. <https://www.gdx.net/core/sample-reports/Melatonin-Sample-Report.pdf>

Genova. (2022c). Menopause Plus™ <https://www.gdx.net/core/sample-reports/Menopause-Plus-Sample-Report.pdf>

Genova. (2022d). Rhythm™. <https://www.gdx.net/core/sample-reports/Rhythm-Sample-Report.pdf>

- Genova_Diagnostics. (2020). *Menopause™ The Original Genova Salivary Sex-Hormone Test*. <https://www.gdx.net/core/sample-reports/Menopause-Sample-Report.pdf>
- Goodman, N. F., Cobin, R. H., Ginzburg, S. B., Katz, I. A., & Woode, D. E. (2011). American Association of Clinical Endocrinologists Medical Guidelines for Clinical Practice for the diagnosis and treatment of menopause. *Endocr Pract*, 17 Suppl 6, 1-25. <https://doi.org/10.4158/EP.17.S6.1>
- Hardiman, P., Thomas, M., Osgood, V., Vlassopoulou, V., & Ginsburg, J. (1990). Are estrogen assays essential for monitoring gonadotropin stimulant therapy? *Gynecol Endocrinol*, 4(4), 261-269. <https://doi.org/10.3109/09513599009024980>
- Hinojosa-Amaya, J. M., Varlamov, E. V., McCartney, S., & Fleseriu, M. (2019). Hypercortisolemia Recurrence in Cushing's Disease; a Diagnostic Challenge. *Front Endocrinol (Lausanne)*, 10, 740. <https://doi.org/10.3389/fendo.2019.00740>
- Kim, Y. J., Kim, J. H., Hong, A. R., Park, K. S., Kim, S. W., Shin, C. S., & Kim, S. Y. (2020). Stimulated Salivary Cortisol as a Noninvasive Diagnostic Tool for Adrenal Insufficiency. *Endocrinol Metab (Seoul)*, 35(3), 628-635. <https://doi.org/10.3803/EnM.2020.707>
- Klee, G. G., & Heser, D. W. (2000). Techniques to measure testosterone in the elderly. *Mayo Clin Proc*, 75 Suppl, S19-25. [https://doi.org/10.1016/S0025-6196\(19\)30637-8](https://doi.org/10.1016/S0025-6196(19)30637-8)
- Lacroix, A., Feelders, R. A., Stratakis, C. A., & Nieman, L. K. (2015). Cushing's syndrome. *Lancet*, 386(9996), 913-927. [https://doi.org/10.1016/s0140-6736\(14\)61375-1](https://doi.org/10.1016/s0140-6736(14)61375-1)
- Lewis, J. G., McGill, H., Patton, V. M., & Elder, P. A. (2002). Caution on the use of saliva measurements to monitor absorption of progesterone from transdermal creams in postmenopausal women. *Maturitas*, 41(1), 1-6. [https://doi.org/10.1016/s0378-5122\(01\)00250-x](https://doi.org/10.1016/s0378-5122(01)00250-x)
- Li, X. S., Li, S., & Kellermann, G. (2018). Simultaneous determination of three estrogens in human saliva without derivatization or liquid-liquid extraction for routine testing via miniaturized solid phase extraction with LC-MS/MS detection. *Talanta*, 178, 464-472. <https://doi.org/10.1016/j.talanta.2017.09.062>
- Martin, K., & Barbieri, R. (2022, August 31). *Menopausal hormone therapy: Benefits and risks*. UpToDate. <https://www.uptodate.com/contents/menopausal-hormone-therapy-benefits-and-risks>
- Meulenberg, P. M., Ross, H. A., Swinkels, L. M., & Benraad, T. J. (1987). The effect of oral contraceptives on plasma-free and salivary cortisol and cortisone. *Clin Chim Acta*, 165(2-3), 379-385. [https://doi.org/10.1016/0009-8981\(87\)90183-5](https://doi.org/10.1016/0009-8981(87)90183-5)
- NAMS. (2012). The 2012 hormone therapy position statement of: The North American Menopause Society. *Menopause*, 19(3), 257-271. <https://doi.org/10.1097/gme.0b013e31824b970a>
- NAMS. (2017). The 2017 hormone therapy position statement of The North American Menopause Society. *Menopause: The Journal of the North American Menopause Society*, 24(7), 728-753. <https://doi.org/10.1097/GME.0000000000000921>
- Nieman. (2015). Cushing's syndrome: update on signs, symptoms and biochemical screening. *Eur J Endocrinol*, 173(4), M33-38. <https://doi.org/10.1530/eje-15-0464>
- Nieman, Biller, B. M., Findling, J. W., Newell-Price, J., Savage, M. O., Stewart, P. M., & Montori, V. M. (2008). The diagnosis of Cushing's syndrome: an Endocrine Society Clinical Practice Guideline. *J Clin Endocrinol Metab*, 93(5), 1526-1540. <https://doi.org/10.1210/jc.2008-0125>
- Nieman, L. K. (2019, September 29). *Measurement of cortisol in serum and saliva*. UpToDate. <https://www.uptodate.com/contents/measurement-of-cortisol-in-serum-and-saliva>
- Nieman, L. K. (2022a, November 12). *Causes and pathophysiology of Cushing's syndrome*. <https://www.uptodate.com/contents/causes-and-pathophysiology-of-cushings-syndrome>
- Nieman, L. K. (2022b, May 29). *Establishing the diagnosis of Cushing's syndrome*. <https://www.uptodate.com/contents/establishing-the-diagnosis-of-cushings-syndrome>

- Nunes, M. L., Vattaut, S., Corcuff, J. B., Rault, A., Loiseau, H., Gatta, B., Valli, N., Letenneur, L., & Tabarin, A. (2009). Late-night salivary cortisol for diagnosis of overt and subclinical Cushing's syndrome in hospitalized and ambulatory patients. *J Clin Endocrinol Metab*, 94(2), 456-462. <https://doi.org/10.1210/jc.2008-1542>
- Oldenburg, M., & Jensen, H. J. (2019). Saliva cortisol level as a strain parameter for crews aboard merchant ships. *Chronobiol Int*, 36(7), 1005-1012. <https://doi.org/10.1080/07420528.2019.1604540>
- Quddusi, S., Browne, P., Toivola, B., & Hirsch, I. B. (1998). Cushing syndrome due to surreptitious glucocorticoid administration. *Arch Intern Med*, 158(3), 294-296. <https://doi.org/10.1001/archinte.158.3.294>
- Rossouw, J. E., Anderson, G. L., Prentice, R. L., LaCroix, A. Z., Kooperberg, C., Stefanick, M. L., Jackson, R. D., Beresford, S. A., Howard, B. V., Johnson, K. C., Kotchen, J. M., & Ockene, J. (2002). Risks and benefits of estrogen plus progestin in healthy postmenopausal women: principal results From the Women's Health Initiative randomized controlled trial. *Jama*, 288(3), 321-333. <https://jamanetwork.com/journals/jama/fullarticle/195120>
- Sakkas, D., Howles, C. M., Atkinson, L., Borini, A., Bosch, E. A., Bryce, C., Cattoli, M., Copperman, A. B., de Bantel, A. F., French, B., Gerris, J., Granger, S. W., Grzegorzczuk-Martin, V., Lee, J. A., Levy, M. J., Matin, M. J., Somers, S., Widra, E. A., & Alper, M. M. (2020). A multi-centre international study of salivary hormone oestradiol and progesterone measurements in ART monitoring. *Reprod Biomed Online*. <https://doi.org/10.1016/j.rbmo.2020.10.012>
- Santoro, N., Braunstein, G. D., Butts, C. L., Martin, K. A., McDermott, M., & Pinkerton, J. V. (2016). Compounded Bioidentical Hormones in Endocrinology Practice: An Endocrine Society Scientific Statement. *J Clin Endocrinol Metab*, 101(4), 1318-1343. <https://doi.org/10.1210/jc.2016-1271>
- Schiffer, L., Adaway, J. E., Arlt, W., & Keevil, B. G. (2019). A liquid chromatography-tandem mass spectrometry assay for the profiling of classical and 11-oxygenated androgens in saliva. *Ann Clin Biochem*, 56(5), 564-573. <https://doi.org/10.1177/0004563219847498>
- Spence, K., McKeever, E., Graham, U., Irwin, S., Neely, J., McAlister, C., Courtney, H., Hunter, S., Mullan, K., McCance, D., & McDonnell, M. (2018). *Salivary cortisol determination using the Roche generation II assay* <https://www.endocrine-abstracts.org/ea/0059/ea0059p007>
- Taylor, H. S., & Manson, J. E. (2011). Update in hormone therapy use in menopause. *J Clin Endocrinol Metab*, 96(2), 255-264. <https://doi.org/10.1210/jc.2010-0536>
- Ueland, G. Å., Kellmann, R., Jørstad Davidsen, M., Viste, K., Husebye, E. S., Almås, B., Storr, H. L., Sagen, J. V., Mellgren, G., Júlíusson, P. B., & Methlie, P. (2021). Bedtime Salivary Cortisol as a Screening Test for Cushing Syndrome in Children. *Journal of the Endocrine Society*, 5(5). <https://doi.org/10.1210/jendso/bvab033>
- UnikeyHealth. (2022). *Salivary Hormone Test*. <https://unikeyhealth.com/products/salivary-hormone-test>
- Valassi, E., Franz, H., Brue, T., Feelders, R. A., Netea-Maier, R., Tsagarakis, S., Webb, S. M., Yaneva, M., Reincke, M., Droste, M., Komerduş, I., Maiter, D., Kastelan, D., Chanson, P., Pfeifer, M., Strasburger, C. J., Toth, M., Chabre, O., Tabarin, A., . . . Trainer, P. J. (2017). Diagnostic tests for Cushing's syndrome differ from published guidelines: data from ERCUSYN. *Eur J Endocrinol*, 176(5), 613-624. <https://doi.org/10.1530/eje-16-0967>
- Wood, P. (2009). Salivary steroid assays - research or routine? *Ann Clin Biochem*, 46(Pt 3), 183-196. <https://doi.org/10.1258/acb.2008.008208>
- Wren, B. G., McFarland, K., Edwards, L., O'Shea, P., Sufi, S., Gross, B., & Eden, J. A. (2000). Effect of sequential transdermal progesterone cream on endometrium, bleeding pattern, and plasma progesterone and salivary progesterone levels in postmenopausal women. *Climacteric*, 3(3), 155-160. <https://doi.org/10.1080/13697130008500109>

Wurtman. (2017). Multiple Sclerosis, Melatonin, and Neurobehavioral Diseases. *Front Endocrinol (Lausanne)*, 8, 280. <https://doi.org/10.3389/fendo.2017.00280>

ZRTLAB. (2019). LCMS Saliva Steroid & Steroid Synthesis Inhibitor Profile. <https://www.zrtlab.com/media/2405/lcms-saliva-steroid-profile-pds.pdf>